



# Republic of the Philippines Department of Health OFFICE OF THE SECRETARY

June 23, 2017

#### **ADMINISTRATIVE ORDER**

No. 2017 - 0010

**SUBJECT: Philippine National Standards for Drinking Water of 2017** 

#### I. RATIONALE

The history of the Philippine National Standards for Drinking Water (PNSDW) started in the year 1963. It was based on the 1958 World Health Organization International Standard for Drinking Water and the 1962 United States Public Health Service Standards. The 1963 PNSDW edition was subsequently revised in 1978, 1993 and 2007.

Since the last revision of PNSDW in 2007, a number of issues and concerns from various stakeholders have emerged. Among these are: (i) experiences of water service providers in complying with the standards; (ii) publication of the fourth edition of the Guidelines for Drinking-Water Quality by the World Health Organization in 2011, which includes new parameters and an improved framework for drinking-water safety that should be considered in water quality monitoring, testing, and analysis; (iii) issuance of DOH Administrative Order Number 2014-0027, which requires all drinking-water service providers to develop and implement water safety plans; (iv) new scope and definitions of Sustainable Development Goal (SDG) water supply indicators; and (v) the need for water quality standards during emergency situations.

This led to the updating of the PNSDW of 2007 through the Inter-Agency Technical Working Group (TWG), headed by the Department of Health (DOH) with support from the World Health Organization (WHO).

#### II. OBJECTIVES

This Administrative Order shall prescribe the standards and procedures on drinking-water quality to protect public/consumer's health.

### III. SCOPE AND COVERAGE

The PNSDW of 2017 shall apply to all drinking-water service providers including government and private developers and operators, bulk water suppliers, water refilling station operators, and water vending machine operators; ice manufacturers; all food establishments, residential, commercial, industrial and institutional buildings that use/supply/serve drinking water; water testing laboratories; health and sanitation authorities; the general public and all others who are involved in determining the safety of public's drinking-water.

#### IV. DEFINITION OF TERMS

- 1. **Acceptability** physical and chemical quality of water that refers to the appearance, taste and odor of drinking-water satisfactory to the consumer.
- 2. **Bulk Water Supply** drinking-water supplied to water service providers or associated infrastructures including pumping stations, reservoirs, and pipe lines.
- 3. **Certified sampling personnel** a person who underwent training for drinking-water sampling and certified by the DOH.
- 4. **Contamination** a general term referring to the presence of substances found in water that make water less desirable or unfit for drinking.
- 5. **Drinking-water** water intended for direct human consumption or for use in food preparation and related processes.
- 6. **Emergency** any situation in which there is actual disruption or damage to communities, i.e., any actual threat to public health and safety.
- 7. **Health-based targets** are measurable health, water quality or performance objectives that are established based on a judgement of safety and on risk assessments of waterborne hazards.
- 8. **Limit of Quantitation** (**LOQ**) the analyte concentration that produces a signal sufficiently stronger than the blank, such that it can be detected with a specified level of reliability during routine operations. Typically, it is the concentration that produces the signal above the reagent water blank signal, and should have a defined precision and bias at that level.
- 9. **Maximum Allowable Level (MAL)** the highest level of a contaminant that is allowed in drinking-water.
- 10. **Method Detection Limit (MDL)** the constituent/contaminant concentration that when processed through the complete method, produces a signal with a 99% probability that is different from the blank.
- 11. **Mobile Water Tanks** tanks designed to deliver water for domestic use or emergency purposes.
- 12. **Potable/Safe Water** water with quality within the standard limits set in this PNSDW both for acceptability and health aspects.
- 13. **Surveillance** the continuous and vigilant public health assessment and review of safety and acceptability of drinking-water supplies.

#### V. GENERAL GUIDELINES

- 1. Standards for drinking-water quality, water sampling and examination and evaluation of results shall conform to the criteria prescribed under this Order and its Manual of Operations.
- 2. To ensure the safety of drinking-water, the standards shall be applied in accordance to the improved framework for drinking-water safety comprising of three key components:
  - A. Health-based targets established by the health authority;
  - B. Safely managed water systems (application of water safety plan); and
  - C. A system of independent surveillance.

#### VI. SPECIFIC GUIDELINES

The Philippine National Standards for Drinking Water of 2017 shall consist of the following criteria:

## 1. Standards for Drinking-water Quality

- A. Drinking-water must be clear and does not have objectionable taste, odor and color. It must be pleasant to drink and free from all harmful organisms, chemical substances and radionuclides in amounts which could constitute a hazard to the health of the consumer.
- B. The quality of drinking-water shall be measured in terms of its microbiological, physical, chemical and radiological constituents. Refer to *Annex A* for the Standard Values and Methods of Analysis.
- C. The parameters of drinking-water quality shall be classified as mandatory, primary and secondary. Refer to *Annex B*.

## 2. Standards for Water Sampling and Examination

- A. Initial examination shall be conducted for new or newly constructed water sources while periodic examination shall be done for existing water sources. Water samples for initial and periodic examination from all water sources shall cover microbiological, physical, chemical and radiological parameters. Refer to *Annex C* for the Minimum Frequency of Sampling.
- B. The minimum number of samples to be collected and examined periodically shall be based on the source and mode of distribution of drinking-water supply. Refer to *Annex C*.
- C. The collection of water samples shall comply with the standard sampling requirements. Refer to  $Annex\ D$ .
- D. Only certified sampling personnel shall collect water samples for regulatory purposes.
- E. All water samples for regulatory purposes shall be examined only in DOH-Accredited Laboratory. The standard methods of examination shall be based on the "22<sup>nd</sup> edition (2012) of the Standard Methods for the Examination of Water and Wastewater" unless otherwise stated in the Manual of Operations.
- F. Examination of water samples for radiological quality shall be done by the Philippine Nuclear Research Institute.

# 3. Standards for Other Modes of Distribution of Drinking-water

- A. Drinking-water from refilling stations, vending machines, mobile tanks and bulk water supply shall be subject for initial and periodic examinations for microbiological, physical, chemical and radiological quality.
- B. All standard values of mandatory parameters shall be applicable to product water from refilling stations and vending machines, except for the standard values of *pH* and total dissolved solids (TDS). The *pH* value shall be 5-7 while the TDS levels of product water shall not exceed 10 mg/L to validate the efficiency of reverse osmosis or distillation process.
- C. Water from mobile tanks shall have chlorine residual (as free chlorine) of at least 0.5 mg/L but not to exceed to 1.50 mg/L at the point of delivery.

- D. Bulk water supply shall maintain chlorine residual (as free chlorine) level between 0.3 mg/L to 1.5 mg/L or chlorine dioxide residual between 0.2 mg/L to 0.4 mg/L prior to distribution.
- E. All water-refilling stations, vending machines, mobile tanks and bulk water supply shall comply with the standard minimum number of samples and frequency of sampling requirements. Refer to *Annex C*.

#### 4. Evaluation of Results

#### A. Expression of Results

- a. Microbiological examination for drinking water shall provide the numbers/presence of Total Coliform, *E. coli*/Thermotolerant Coliform, and Heterotrophic Bacteria present in 100 mL of water.
- b. All results from physico-chemical and radiological examinations that are not detected shall be reported as less than the method detection limit (MDL). For trace analysis, the MDL and level of quantitation (LOQ) shall be reflected in the laboratory test report.

## B. Interpretation of Results

- a. Drinking-water service providers shall consult the DOH/Local Drinking Water Quality Monitoring Committee (LDWQMC)/local health office for the interpretation of results.
- b. When *E. coli*/Thermotolerant Coliform is present in water, a sanitary survey shall be conducted within 24 hours to determine the cause of contamination which include resampling. If resampled water still contains *E. coli*/Thermotolerant coliform, corrective actions should be applied. At the same time, the drinking-water service provider shall issue an advisory to "boil water" or other household water treatment options, or provide an alternative drinking-water supply.
- c. In case of exceedance of standard values of physical and chemical parameters, monitoring shall be carried out for the next three (3) consecutive months wherein all results must comply with the standards. If the results still exceed, further study must be done to determine the cause of contamination for proper identification of corrective actions.

### 5. Classification of Drinking-Water Quality Parameters

### A. Mandatory Parameters

a. Mandatory parameters are legally enforceable. These core parameters shall be required for examination by all drinking-water service providers. The criteria used for selection of mandatory core parameters are: parameters that directly affect health through acute or chronic exposure and/or will render the water unacceptable for drinking; indicate the possible presence of other contaminants; exceed tolerable values/standards based on local monitoring data of the previous years; have wide spatial distribution across the Philippines based on local monitoring data; and viable indicators for general quality and stability of water supply.

- b. The frequency of testing for mandatory parameters, except for *E*. coli/Thermotolerant Coliform and residual disinfectant, may be reduced to every three (3) years if the LDWQMC found the consolidated water quality reports showed undetectable levels (below MDL) of a particular mandatory parameter for three (3) consecutive years.
- c. The mandatory parameters are the minimum parameters required to be tested for initial and periodic examinations. However, the mandatory parameters may include additional parameters from the list of primary and secondary as determined by the LDWQMC. The additional parameters shall be based on the result of the risk assessment of the water sources where potential contamination from the natural or anthropogenic activities may occur.

# B. Primary parameters

- a. Primary parameters are site-specific. These are chemical impurities in water that directly affect health through acute or chronic exposure.
- b. Primary parameters can also be adopted as enforceable parameters, in addition to the mandatory parameters.

## C. Secondary parameters

- a. Secondary parameters are those that render the water unacceptable for drinking.
- b. These include operational parameters which affect the efficiency of the treatment processes.

## D. Emergency Drinking-Water Parameters

- a. During the first 72 hours, temporary supply of water shall be provided by the local government unit (LGU). Water should be disinfected as a minimum treatment (i.e. boiling, chlorination, etc.). Mobile treatment plant can be used as an alternative source of water.
- b. The water supply shall be monitored daily for at least seven (7) days by the LGU and other respondents in terms of residual chlorine, and *E. coli*. The acceptable level of residual chlorine shall be 0.5 mg/L and a maximum level of 1.5 mg/L. *E. coli* should be absent per 100 mL sample.
- c. Regular monitoring shall resume after normal condition has been declared by the appropriate government agency.

#### E. Sustainable Development Goal (SDG) Parameters

- a. Relative to Target 6.1.1 of the SDG in achieving universal and equitable access to safe and affordable drinking water for all by 2030, the population should be using safely managed drinking water services.
- b. This entails that the population uses a drinking water source which is located on premises, available when needed, and free of fecal and priority chemical contamination.

## 6. Quality Assurance/Quality Control for Water Laboratories

- A. Only laboratories accredited by the DOH shall perform drinking-water quality examination for regulatory purposes.
- B. All accredited laboratories shall provide highest quality service through the establishment, documentation, and effective operation of a Quality System (QS).
- C. The laboratory personnel involved in water sampling shall be certified by the DOH.

## 7. Water Safety Plan (WSP) and Drinking-water Quality Surveillance

- A. The implementation of WSP approach can secure the safety of drinking-water. It utilizes a risk assessment and risk management approach that encompasses all steps in the water supply system, from catchment/source to consumers.
- B. All drinking-water service providers shall be required to prepare WSP as provided by the DOH Administrative Order No. 2014 0027 "National Policy on Water Safety Plan for All Drinking-Water Service Providers", dated September 4, 2014.
- C. The WSP of a drinking-water service provider shall be subject for review and approval as provided by the DOH Administrative Order No. 2017-0006 "Guidelines for the Review and Approval of the Water Safety Plans of Drinking-Water Service Providers", dated April 20, 2017.
- D. The WSP shall be developed to meet health-based targets consistent with the Philippine National Standards for Drinking Water.
- E. The drinking-water quality surveillance agency shall ensure that monitoring of the WSP implementation and its effectiveness meets the Philippine National Standards for Drinking Water. The surveillance activity shall include audit and direct assessment approaches.

#### VII. ROLES AND RESPONSIBILITIES

### A. Department of Health

- a. Develop systems and procedures to operationalize this Order.
- b. Ensure compliance of all drinking-water service providers and operators to this Order.
- c. Perform independent surveillance of drinking-water service providers.
- d. Provide technical assistance to the local government units, drinking-water service providers and to the general public.
- e. Accredit water laboratories, certify training providers and water sampling personnel.

#### B. Local Government Unit

- a. Enforce the provisions of this Order.
- b. Develop and implement drinking water quality surveillance program.
- c. Establish a local drinking water quality monitoring committee.
- d. Advocate and create awareness to general population on the importance of drinking water quality standards, impact of water contamination on health, and control measures on addressing water quality issues and problems.

### C. Water Laboratory

- a. Comply with the provisions of this Order.
- b. Secure accreditation from the Department of Health.
- c. Implement QS and develop a manual of operations describing the laboratory's policies and plans for ensuring the quality of their work provided to the public.

## D. Drinking-Water Service Provider/Operator of Establishment and Building

- a. Comply with the provisions of this Order.
- b. Develop and implement WSP.
- c. Institute corrective actions for any unsatisfactory results of water sampling.
- d. Submit to the accredited laboratories water samples for examination in a manner and at such intervals prescribed under this Order.
- e. Submit results of water quality testing to the local health authority.
- f. Educate consumers on how to keep drinking-water safe at all times.

#### VIII. PENAL PROVISION

As provided in Sec. 103 of the Code on Sanitation of the Philippines (PD No. 856):

- A. Any person who shall violate, disobey, refuse, omit or neglect to comply with any of the provisions of this Order, shall be guilty of misdemeanor and upon conviction shall be punished by imprisonment for a period of not exceeding six (6) months or by a fine of not exceeding Php 1,000.00 or both depending upon the discretion of the court.
- B. Any person who shall interfere with or hinder, or oppose any officer, agent or member of the Department or of the bureaus and offices under it, in the performance of his duty as such under this Order, or shall tear down, mutilate, deface or alter any placard, or notice, affixed to the premises in the enforcement of this Order, shall be guilty of a misdemeanor and punishable upon conviction by imprisonment for a period of not exceeding six (6) months or by a fine of not exceeding Php 1,000.00 or both depending upon the discretion of the court.

#### IX. SEPARABILITY CLAUSE

In the event that any rule, section, paragraph, sentence, clause, or word of this Order is declared invalid for any reason, the other provisions thereof shall not be affected thereby.

# X. REPEALING CLAUSE

Administrative Order No. 2007 - 0012 (2007 PNSDW) is hereby repealed. All laws, rules and regulations and administrative issuances or parts thereof inconsistent with the provisions of these standards are hereby repealed or amended accordingly.

## XI. EFFECTIVITY

This order takes effect fifteen (15) days after its publication in an official gazette or in a newspaper of general circulation.

PAULYN JEAN B. ROSELL-UBIAL, MD, MPH, CESO II

Secretary of Health

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Table A-1. Standard Values, Methods of Detection and Points of Compliance for Microbiological Quality of Drinking-Water

| Parameter                                | Standard Values   | Methods of Analysis (SMEWW 22 <sup>nd</sup> ed.)  | Point of Compliance   |  |
|--|---|---|---|--|
| 1. Total Coliform                        | MTFT:<br>< 1.1 MPN/ 100<br>mL                               | • 9221 Multiple Tube<br>Fermentation Technique  | <ul><li>Consumer's taps</li><li>Water treatment<br/>works/plants</li></ul>  |  |
|  | EST:<br>Absent or <1<br>MPN/100 mL                          | • 9223 Enzyme Substrate<br>Coliform Test*   | <ul> <li>Water refilling stations</li> <li>Water vending machines</li> <li>Mobile treatment devices</li> </ul>  |  |
|  | MFT:<br>< 1 total coliform<br>colonies / 100 mL             | <ul> <li>9222B Standard Total         Coliform Membrane Filter         Technique</li> <li>9222CDelayed Incubation         Total Coliform Procedure*</li> <li>9222H Simultaneous         Detection of Total         Coliform and E. coli by         Dual-Chromogen         Membrane Filter         Technique*</li> </ul> | <ul> <li>Point of use treatment devices</li> <li>Water haulers</li> <li>Bulk Water</li> </ul>   |  |
| 2. Thermotolerant<br>Coliform/E.coli     | MTFT:<br>< 1.1 MPN/ 100<br>mL                               | <ul> <li>9221 Multiple Tube         Fermentation Technique</li> <li>9221 E1 Thermotolerant         Coliform Test (EC         medium)</li> <li>9221 E2 Thermotolerant         Coliform Test (A-1         medium)*</li> </ul>   | <ul> <li>Point sources</li> <li>Consumer's taps</li> <li>Water treatment works</li> <li>Water refilling stations</li> <li>Water vending machines</li> <li>Mobile treatment devices</li> <li>Point of use treatment devices</li> </ul>   |  |
|  | EST:<br>Absent or <1<br>MPN/100 mL                          | • 9223 Enzyme Substrate<br>Coliform Test*   | <ul><li>Water haulers</li><li>Bulk Water</li><li>Food Establishments</li></ul>  |  |
|  | MFT:<br>< 1 thermotolerant<br>coliform colonies /<br>100 mL | • 9222B Standard Total<br>Coliform Membrane Filter<br>Technique   | <ul><li> All buildings</li><li> Ice Plants</li></ul>  |  |
| 3. Heterotrophic<br>Plate Count<br>(HPC) | <500 CFU/mL   | <ul> <li>9215 B Pour Plate Method</li> <li>9215 C Spread Plate<br/>Method</li> <li>9215 D Membrane Filter<br/>Method</li> </ul>   | <ul> <li>Consumer's taps</li> <li>Water treatment works</li> <li>Water refilling stations</li> <li>Water vending machines</li> <li>Mobile treatment devices</li> <li>Point of use treatment devices</li> <li>Water haulers</li> <li>Bulk Water</li> <li>Food Establishments</li> <li>All buildings</li> <li>Ice Plants</li> </ul> |  |

MTFT: Multiple Tube Fermentation Technique, MPN: Most Probable Number EST: Enzyme Substrate Test, CFU: Colony Forming Units MFT: Membrane Filter Technique,\*should be verified and approved by the DOH

Table A-2. Summary of Standard Values and Methods of Analysis for Inorganic Chemical Parameters of Drinking-Water

| Parameter        | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level (MAL) | Methods of Analysis   |
|------------------|--|-------------------------------------|---|
| 1. Antimony (Sb) | 7440-36-0  | 0.02 mg/L                           | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 F. Nitric Acid-Hydrochloric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3113 B. Electrothermal Atomic Absorption Spectrometric Method</li> <li>3120 B. Inductively Coupled Plasma Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry</li> </ul>   |
| 2. Arsenic(As)   | 7440-38-2  | 0.01 mg/L                           | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 G. Nitric Acid-Sulfuric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3114 B. Manual Hydride Generation/Atomic Absorption Spectrometric Method</li> <li>3113 B. Electrothermal Atomic Absorption Spectrometric</li> <li>3120 B. Inductively Coupled Plasma Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry Method</li> </ul> |
| 3. Barium (Ba)   | 7440-39-3  | 0.70 mg/L                           | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 F. Nitric Acid-Hydrochloric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3111 D. Direct Nitrous Oxide-Acetylene Flame Method</li> <li>3113 B. Electrothermal Atomic Absorption Spectrometric</li> <li>3120 B. Inductively Coupled Plasma Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry Method</li> </ul>                  |
| 4. Boron (B)     | 7440-42-8  | 2.00 mg/L                           | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 G. Nitric Acid-Sulfuric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3120 B. Inductively Coupled Plasma Method</li> </ul>  |

| Parameter                               | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level (MAL) | Methods of Analysis  |
|---|--|-------------------------------------|--|
|   |  |                                     | 3125 B. Inductively Coupled Plasma-Mass<br>Spectrometry Method     4500B B. Curcumin Method     4500B C. Carmine Method  |
| 5. Cadmium<br>(Cd)                      | 7440-43-9  | 0.003 mg/L                          | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 F. Nitric Acid-Hydrochloric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3113 B. Electrothermal Atomic Absorption Spectrometric</li> <li>3120 B. Inductively Coupled Plasma Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry Method</li> </ul>        |
| 6. Chromium,<br>Total (Cr)              | 7440-47-3  | 0.05 mg/L                           | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 F. Nitric Acid-Hydrochloric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3113 B. Electrothermal Atomic Absorption Spectrometric Method</li> <li>3120 B. Inductively Coupled Plasma Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry Method</li> </ul> |
| 7. Cyanide,<br>Total (CN <sup>-</sup> ) | 57-12-5  | 0.50 mg/L                           | <ul> <li>4500-CN<sup>-</sup> D. Titrimetric Method</li> <li>4500-CN<sup>-</sup> E. Colorimetric Method</li> <li>4500-CN<sup>-</sup> F. Cyanide-Selective Electrode<br/>Method</li> </ul>   |
| 8. Fluoride (F <sup>-</sup> )           | 16984-48-<br>8                                   | 1.50 mg/L                           | A. Sample Preparation Preliminary Distillation B. Instrumentation  4110 B. Ion Chromatography with Chemical Suppression of Eluent Conductivity  4110 C. Single-Column Ion Chromatography with Direct Conductivity Detection  4500-F C. Ion-selective Electrode Method  |
| 9. Lead (Pb)                            | 7439-92-1  | 0.01 mg/L                           | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3113 B. Electrothermal Atomic Absorption<br/>Spectrometric</li> <li>3120 B. Inductively Coupled Plasma<br/>Method</li> </ul>  |

| Parameter                                   | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level (MAL) | Methods of Analysis   |
|---|--|-------------------------------------|---|
|   |  |                                     | · 3125 B. Inductively Coupled Plasma-Mass<br>Spectrometry Method  |
| 10. Manganese<br>(Mn)                       | N/A  | 0.4 mg/L                            | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 F. Nitric Acid-Hydrochloric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3111 B. Direct Air-Acetylene Flame Method</li> <li>3111 C. Extraction/Air-Acetylene Flame Method</li> <li>3113 B. Electrothermal Atomic Absorption Spectrometric</li> <li>3120 B. Inductively Coupled Plasma Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry Method</li> </ul>           |
| 11. Mercury,<br>Total (Hg)                  | 7439-97-6  | 0.001 mg/L                          | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 G. Nitric Acid-Sulfuric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3112 B. Cold-Vapor Atomic Absorption Spectrometric Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry Method</li> <li>EPA 245.7 / BS EN 13506 Atomic Fluorescence Spectrometric Method</li> <li>EPA 7473 Thermal Decomposition, Amalgamation, Atomic Absorption Spectrometric Method</li> </ul> |
| 12. Nickel (Ni)                             | 7440-02-0  | 0.07 mg/L                           | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 F. Nitric Acid-Hydrochloric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3113 B. Electrothermal Atomic Absorption Spectrometric</li> <li>3120 B. Inductively Coupled Plasma Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry Method</li> </ul>   |
| 13. Nitrate (NO <sub>3</sub> <sup>-</sup> ) | C-005  | 50.00 mg/L                          | <ul> <li>4110 B. Ion Chromatography with<br/>Chemical Suppression of Eluent<br/>Conductivity</li> <li>4110 C. Single-Column Ion<br/>Chromatography with Direct Conductivity<br/>Detection</li> </ul>  |

| Parameter                      | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level (MAL) | Methods of Analysis  |
|--------------------------------|--|-------------------------------------|--|
|                                |  |                                     | <ul> <li>4500-NO<sub>3</sub><sup>-</sup> B. Ultraviolet         Spectrophotometric Screening Method</li> <li>4500-NO<sub>3</sub><sup>-</sup> E. Cadmium Reduction Method</li> <li>4500-NO<sub>3</sub><sup>-</sup> I. Cadmium Reduction Flow Injection Method</li> <li>4140. Capillary Ion electrophoresis</li> <li>4500-NO<sub>3</sub><sup>-</sup> D. Nitrate Electrode Method</li> </ul>  |
| 14.Nitrite (NO <sub>2</sub> -) | C-005  | 3.00 mg/L                           | <ul> <li>4110 B. Ion Chromatography with<br/>Chemical Suppression of Eluent<br/>Conductivity</li> <li>4110 C. Single-Column Ion<br/>Chromatography with Direct Conductivity<br/>Detection</li> <li>4500-NO<sub>2</sub>- B. Colorimetric Method</li> <li>4130. Flow Injection Analysis</li> <li>4140. Capillary Ion electrophoresis</li> <li>4500-NO<sub>2</sub>- B. Colorimetric (Diazotization)</li> </ul>  |
| 15.Selenium (Se)               | 7782-49-2  | 0.04 mg/L                           | <ul> <li>A. Sample Preparation</li> <li>3030 E. Nitric Acid Digestion</li> <li>3030 G. Nitric Acid-Sulfuric Acid Digestion</li> <li>3030 K. Microwave-Assisted Digestion</li> <li>B. Instrumentation</li> <li>3114 B. Manual Hydride Generation/Atomic Absorption Spectrometric Method</li> <li>3114 C. Continuous Hydride Generation AAS</li> <li>3113 B. Electrothermal Atomic Absorption Spectrometric</li> <li>3120 B. Inductively Coupled Plasma Method</li> <li>3125 B. Inductively Coupled Plasma-Mass Spectrometry Method</li> </ul> |

Table A-3. Summary of Standard Values and Methods of Analysis for Organic Chemical Parameters from Industrial Pollution of Drinking-Water

|     | Parameter                     | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level<br>(MAL)<br>(mg/L) | Method of Analysis   |
|-----|-------------------------------|--|--|--|
| 1.  | Benzene                       | 71-43-2  | 0.01   | <ul> <li>6200B. Purge and Trap Capillary-Column<br/>Gas Chromatographic/Mass Spectrometric<br/>Method</li> <li>6200B. Purge and Trap Capillary-Column<br/>Gas Chromatographic/Mass Spectrometric<br/>Method</li> <li>6200 C. Purge and Trap Capillary Column<br/>Gas Chromatographic Method</li> </ul> |
| 2.  | Benzo(a)pyrene                | 50-32-8  | 0.0007   | 6410B. Liquid-Liquid Extraction Gas<br>Chromatographic/Mass Spectrometric<br>Method     6440B. Liquid-Liquid Chromatographic<br>Method   |
| 3.  | Carbon Tetrachloride          | 56-23-5  | 0.004  | <ul> <li>6200B. Purge and Trap Capillary-Column<br/>Gas Chromatographic/Mass Spectrometric<br/>Method</li> <li>6200 C. Purge and Trap Capillary Column<br/>Gas Chromatographic Method</li> </ul>   |
| 4.  | 1,2-Dichlorobenzene           | 95-50-1  | 1  | <ul> <li>6200B. Purge and Trap Capillary-Column<br/>Gas Chromatographic/Mass Spectrometric<br/>Method</li> <li>6200 C. Purge and Trap Capillary Column<br/>Gas Chromatographic Method</li> </ul>   |
| 5.  | 1,4-Dichlorobenzene           | 106-46-7   | 0.3  | <ul> <li>6200B. Purge and Trap Capillary-Column<br/>Gas Chromatographic/Mass Spectrometric<br/>Method</li> <li>6200 C. Purge and Trap Capillary Column<br/>Gas Chromatographic Method</li> </ul>   |
| 6.  | 1,2-Dichloroethane            | 107-06-2   | 0.03   | <ul> <li>6200B. Purge and Trap Capillary-Column<br/>Gas Chromatographic/Mass Spectrometric<br/>Method</li> <li>6200 C. Purge and Trap Capillary Column<br/>Gas Chromatographic Method</li> </ul>   |
| 7.  | 1,2-Dichloroethene            | 156-59-2<br>(cis)<br>156-60-5<br>(trans)         | 0.05   | 6200B. Purge and Trap Capillary-Column     Gas Chromatographic/Mass Spectrometric     Method     6200 C. Purge and Trap Capillary Column     Gas Chromatographic Method  |
| 8.  | Dichloromethane               | 75-09-2  | 0.02   | <ul> <li>6200B. Purge and Trap Capillary-Column<br/>Gas Chromatographic/Mass Spectrometric<br/>Method</li> <li>6200 C. Purge and Trap Capillary Column<br/>Gas Chromatographic Method</li> </ul>   |
| 9.  | Di(2-ethylhexyl)<br>phthalate | 117-81-7   | 0.008  | 6410B. Liquid-Liquid Extraction Gas<br>Chromatographic/Mass Spectrometric<br>Method  |
| 10. | Ethylbenzene                  | 100-41-4   | 0.3  | <ul> <li>6200B. Purge and Trap Capillary-Column<br/>Gas Chromatographic/Mass Spectrometric<br/>Method</li> <li>6200 C. Purge and Trap Capillary Column<br/>Gas Chromatographic Method</li> </ul>   |

| Parameter             | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level<br>(MAL)<br>(mg/L) | Method of Analysis  |
|-----------------------|--|--|---|
| 11. Styrene           | 100-42-5   | 0.02   | 6200B. Purge and Trap Capillary-Column     Gas Chromatographic/Mass Spectrometric     Method     6200 C. Purge and Trap Capillary Column     Gas Chromatographic Method |
| 12. Tetrachloroethene | 127-18-4   | 0.04   | 6200B. Purge and Trap Capillary-Column Gas Chromatographic/Mass Spectrometric Method     6200 C. Purge and Trap Capillary Column Gas Chromatographic Method             |
| 13. Toluene           | 108-88-3   | 0.7  | 6200B. Purge and Trap Capillary-Column     Gas Chromatographic/Mass Spectrometric     Method     6200 C. Purge and Trap Capillary Column     Gas Chromatographic Method |
| 14. Vinyl Chloride    | 75-01-4  | 0.0003   | 6200B. Purge and Trap Capillary-Column Gas Chromatographic/Mass Spectrometric Method     6200 C. Purge and Trap Capillary Column Gas Chromatographic Method             |
| 15. Xylenes (total)   | 1330-20-7  | 0.5  | 6200B. Purge and Trap Capillary-Column Gas Chromatographic/Mass Spectrometric Method     6200 C. Purge and Trap Capillary Column Gas Chromatographic Method             |

Table A-4. Summary of Standard Values and Methods of Analysis for Organic Chemical Parameters (Pesticides) of Drinking-Water

|    | Parameter                             | Chemical             | Maximu         | Method of Analysis   |
|----|---------------------------------------|----------------------|----------------|--|
|    |                                       | Abstracts<br>Service | m<br>Allowable |  |
|    |                                       | (CAS)                | Level          |  |
|    |                                       | No.                  | (mg/L)         |  |
| 1. | Aldrin and Dieldrin                   | Aldrin:              | 0.00003        | 6410B. Liquid-Liquid Extraction  |
|    | (combined)                            | 309-00-2             |                | Gas Chromatographic/Mass   |
|    |                                       | Dieldrin:            |                | Spectrometric Method   |
|    |                                       | 60-57-1              |                | 6630B. Liquid-Liquid Extraction     Gas Chromatographic Method I               |
|    |                                       |                      |                | • 6630C. Liquid-Liquid Extraction  |
|    |                                       |                      |                | Gas Chromatographic Method II  |
| 2. | Atrazine and its chloro-s-triazine    | 1912-24-9            | 0.1            | • USEPA 525.2 Gas  |
|    | metabolites                           |                      |                | Chromatography/Mass Spectrometry   |
|    |                                       |                      |                | 6630C. Liquid-Liquid Extraction     Con Classical Methods                      |
|    |                                       |                      |                | Gas Chromatographic Method II  |
| 3. | Carbofuran                            | 1563-66-2            | 0.007          | 6610B. High-Performance Liquid<br>Chromatographic Method                       |
| 4. | Chlordane                             | 57-74-9              | 0.0002         | 6410B. Liquid-Liquid Extraction  |
|    |                                       |                      |                | Gas Chromatographic/Mass   |
|    |                                       |                      |                | <ul><li>Spectrometric Method</li><li>6630B. Liquid-Liquid Extraction</li></ul> |
|    |                                       |                      |                | Gas Chromatographic Method I   |
|    |                                       |                      |                | • 630C. Liquid-Liquid Extraction Gas   |
|    |                                       |                      |                | Chromatographic Method II  |
| 5. | 1,2-Dibromo-3-chloropropane           | 96-12-8              | 0.001          | • 6200B. Purge and Trap Capillary-   |
|    | (DBCP)                                |                      |                | Column Gas Chromatographic/Mass<br>Spectrometric Method                        |
|    |                                       |                      |                | • 6200 C. Purge and Trap Capillary   |
|    |                                       |                      |                | Column Gas Chromatographic   |
|    |                                       |                      |                | Method   |
|    |                                       |                      |                | • 6231B Liquid-liquid extraction- Gas  |
| 6  | Dichloredinhanvitrichloreethone       | 50-29-3              | 0.001          | Chromatographic Method   |
| 6. | Dichlorodiphenyltrichloroethane (DDT) | 30-29-3              | 0.001          | 6410B. Liquid-Liquid Extraction     Gas Chromatographic/Mass                   |
|    | (DD1)                                 |                      |                | Spectrometric Method   |
|    |                                       |                      |                | 6630B. Liquid-Liquid Extraction  |
|    |                                       |                      |                | Gas Chromatographic Method I   |
|    |                                       |                      |                | 6630C. Liquid-Liquid Extraction  |
| 7. | Endrin                                | 72-20-8              | 0.0006         | Gas Chromatographic Method II  |
| '. | Endin                                 | 12-20-8              | 0.0006         | 6410B. Liquid-Liquid Extraction     Gas Chromatographic/Mass                   |
|    |                                       |                      |                | Spectrometric Method   |
|    |                                       |                      |                | 6630B. Liquid-Liquid Extraction  |
|    |                                       |                      |                | Gas Chromatographic Method I   |
|    |                                       |                      |                | • 6630C. Liquid-Liquid Extraction  |
|    |                                       |                      |                | Gas Chromatographic Method II  |
| 8. | Ethylene Dibromide or 1,2-            | 106-93-4             | 0.0004         | • 6200B. Purge and Trap Capillary-   |
|    | Dibromoethane                         |                      |                | Column Gas Chromatographic/Mass  |
|    |                                       |                      |                | Spectrometric Method   |
|    |                                       |                      |                | • 6200 C. Purge and Trap Capillary   |
|    |                                       |                      |                | Column Gas Chromatographic Method  |
| 9. | Glyphosate                            | 1071-83-6            | 1              | 6651B. Liquid Chromatographic  |
|    | ••                                    |                      |                | Post-Column Fluorescence Method  |

| Parameter         | Chemical  | Maximu    | Method of Analysis                                  |
|-------------------|-----------|-----------|---|
|                   | Abstracts | m         |   |
|                   | Service   | Allowable |   |
|                   | (CAS)     | Level     |   |
|                   | No.       | (mg/L)    |   |
| 10. Lindane       | 58-89-9   | 0.002     | 6410B. Liquid-Liquid Extraction                     |
|                   |           |           | Gas Chromatographic/Mass                            |
|                   |           |           | Spectrometric Method                                |
|                   |           |           | <ul> <li>6630B. Liquid-Liquid Extraction</li> </ul> |
|                   |           |           | Gas Chromatographic Method I                        |
|                   |           |           | 6630C. Liquid-Liquid Extraction Gas                 |
|                   |           |           | Chromatographic Method II                           |
| 11. Pendimethalin | 40487-42- | 0.02      | USEPA 525.2 Liquid-solid                            |
|                   | 1         |           | extraction and capillary column Gas                 |
|                   |           |           | Chromatography/Mass Spectrometry                    |

Table A-5. Summary of Standard Values and Methods of Analysis for Physical and Chemical Quality for Acceptability Aspects of Drinking-Water

|    | Parameter                           | Chemical<br>Abstracts<br>Service<br>(CAS) No. | Maximum<br>Allowable<br>Level (MAL) | Methods of Analysis  |
|----|-------------------------------------|---|-------------------------------------|--|
| 1. | Taste                               | N/A   | No<br>objectionable<br>taste        | Sensory Evaluation Technique  Testing of taste shall be based on consumers' complaints.  |
| 2. | Odor                                | N/A   | No<br>objectionable<br>odor         | Sensory Evaluation Technique   |
| 3. | Color (Apparent)                    | N/A   | 10 CU                               | 2120 B. Visual Comparison Method – for apparent color only   |
| 4. | Turbidity                           | N/A   | 5 NTU                               | 2130 B. Nephelometric Method   |
| 5. | Aluminum (Al)                       | N/A   | 0.2 mg/L<br>(Aesthetic)             | A. Sample Preparation 3030 E. Nitric Acid Digestion 3030 F. Nitric Acid-Hydrochloric Acid Digestion 3030 K. Microwave-Assisted Digestion B. Instrumentation 3500-Al B. Eriochrome Cyanine R Method 3113 B. Electrothermal Atomic Absorption Spectrometric 3120 B. Inductively Coupled Plasma Method 3125 B. Inductively Coupled Plasma-Mass Spectrometry Method  |
| 6. | Chloride (Cl <sup>-</sup> )         | 16887-00-6                                    | 250 mg/L                            | <ul> <li>4500 - Cl- B Argentometric method</li> <li>4500 - Cl- D Potentiometric Method</li> <li>4110 B. Ion Chromatography with<br/>Chemical Suppression of Eluent<br/>Conductivity</li> <li>4110 C. Single-Column Ion<br/>Chromatography with Direct Conductivity<br/>Detection</li> </ul>  |
| 7. | Copper (Cu)                         | N/A   | 1.0 mg/L                            | A. Sample Preparation  3030 E. Nitric Acid Digestion  3030 F. Nitric Acid-Hydrochloric Acid Digestion  3030 K. Microwave-Assisted Digestion  B. Instrumentation  3111 B. Direct Air-Acetylene Flame Method  3111 C. Extraction/Air-Acetylene Flame Method  3113 B. Electrothermal Atomic Absorption Spectrometric  3120 B. Inductively Coupled Plasma Method  3125 B. Inductively Coupled Plasma-Mass Spectrometry Method  3500-Cu C. Bathocuproine Method |
| 8. | Total Hardness                      | N/A   | 300 mg/L                            | 2430 C EDTA Titrimetric method   |
| 9. | Hydrogen sulfide (H <sub>2</sub> S) | 7783-06-4                                     | 0.05 mg/L                           | <ul> <li>4500 S<sup>2-</sup> D. Methylene Blue Method</li> <li>4500 S<sup>2-</sup> E. Gas Dialysis, Automated<br/>Methylene Blue Method</li> </ul>   |

|     | Parameter                                | Chemical<br>Abstracts<br>Service<br>(CAS) No. | Maximum<br>Allowable<br>Level (MAL) | Methods of Analysis  |
|-----|--|---|-------------------------------------|--|
|     |  |   |                                     | <ul> <li>4500 S<sup>2-</sup> I. Distillation, Methylene Blue<br/>Flow Injection Analysis Method</li> <li>4500 S<sup>2-</sup> F. Iodometric Method</li> </ul>   |
| 10. | Iron (Fe)                                | N/A   | 1.0 mg/L                            | A. Sample Preparation  3030 E. Nitric Acid Digestion  3030 F. Nitric Acid-Hydrochloric Acid Digestion  3030 K. Microwave-Assisted Digestion  B. Instrumentation  3111 B. Direct Air-Acetylene Flame Method  3111 C. Extraction/Air-Acetylene Flame Method  3113 B. Electrothermal Atomic Absorption Spectrometric  3120 B. Inductively Coupled Plasma Method  3125 B. Inductively Coupled Plasma-Mass Spectrometry Method  3500-Fe B. Phenanthroline Method  |
| 11. | pH                                       | N/A   | 6.5 - 8.5                           | 4500-H <sup>+</sup> B. Electrometric Method  |
| 12. | Sodium (Na)                              | N/A   | 200 mg/L                            | A. Sample Preparation  3030 E. Nitric Acid Digestion 3030 F. Nitric Acid-Hydrochloric Acid Digestion 3030 K. Microwave-Assisted Digestion  B. Instrumentation 3111 B. Direct Air-Acetylene Flame Method 3113 B. Electrothermal Atomic Absorption Spectrometric 3120 B. Inductively Coupled Plasma Method 3125 B. Inductively Coupled Plasma-Mass Spectrometry Method 3500-Na B. Flame Emission Photometric Method  |
| 13. | Sulfate (SO <sub>4</sub> <sup>2-</sup> ) | 14808-79-8                                    | 250 mg/L                            | <ul> <li>4110 B. Ion Chromatography with Chemical Suppression of Eluent Conductivity</li> <li>4140 B. Capillary Ion Electrophoresis with Indirect UV Detection</li> <li>4500-SO<sub>4</sub><sup>2-</sup> C. Gravimetric method with Ignition of Residue</li> <li>4500-SO<sub>4</sub><sup>2-</sup> D. Gravimetric Method with Drying of Residue</li> <li>4500-SO<sub>4</sub><sup>2-</sup> E. Turbidimetric Method</li> <li>4500-SO<sub>4</sub><sup>2-</sup> F. Automated Methylthymol Blue Method</li> <li>4500-SO<sub>4</sub><sup>2-</sup> G. Methylthymol Blue Flow Injection Analysis</li> </ul> |
| 14. | Total Dissolved<br>Solids                | N/A   | 600 mg/L                            | 2540 C. Total Dissolved Solids Dried at 180°C  |

| Parameter     | Chemical<br>Abstracts<br>Service<br>(CAS) No. | Maximum<br>Allowable<br>Level (MAL) | Methods of Analysis  |
|---------------|---|-------------------------------------|--|
| 15. Zinc (Zn) | N/A   | 5.0 mg/L                            | A. Sample Preparation 3030 E. Nitric Acid Digestion 3030 F. Nitric Acid-Hydrochloric Acid Digestion 3030 K. Microwave-Assisted Digestion B. Instrumentation 3111 B. Direct Air-Acetylene Flame Method 3113 B. Electrothermal Atomic Absorption Spectrometric 3111 C. Extraction/Air-Acetylene Flame Method  3120 B. Inductively Coupled Plasma Method 3125 B. Inductively Coupled Plasma-Mass Spectrometry Method 3500-Zn B. Zincon Method |

Table A-6. Summary of Standard Values and Methods of Analysis for Treatment Chemicals
Used in Treatment and Disinfection and Disinfection by-products of Drinking-Water

| Parameter                               | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level (mg/L) | Method of Analysis  |
|---|--|--------------------------------------|---|
| a. Contaminants                         | from Treatme                                     | ent Chemicals                        |   |
| 1. Acrylamide                           | 79-06-1  | 0.0005                               | USEPA 8316 High-performance     Liquid Chromatography with UV     Detection   |
| 2. Epichlorohydrin                      | 106-89-8   | 0.0004                               | 6200 B. Purge and Trap Capillary-<br>Column Gas Chromatographic/Mass<br>Spectrometric Method  |
| b. Disinfection C                       | hemicals   |                                      |   |
| 1. Chlorine Dioxide Residual            | 10049-04-4                                       | 0.2 min and 0.4<br>max <sup>1</sup>  | Colorimeter-Refer to manufacturer's<br>manual provided with the test equipt.  |
| 2. Chlorine Residual (as free chlorine) | Chlorine: 7782-50-5                              | 0.3 min and 1.5<br>max               | DPD Colorimetric Method –Refer to<br>manufacturer's manual provided with<br>the test kit  |
| c. Disinfection By                      |  |                                      |   |
| 1. Bromate                              | 15541-45-4                                       | 0.01                                 | <ul> <li>4110 B. Ion Chromatography with<br/>Chemical Suppression of Eluent<br/>Conductivity</li> <li>4110 C. Single-Column Ion<br/>Chromatography with Direct<br/>Conductivity Detection</li> <li>4110 D. Ion Chromatographic<br/>Determination of Oxyhalides and<br/>Bromide</li> </ul> |
| 2. Chlorate                             | 7775-09-9  | 0.7                                  | 4110 B. Ion Chromatography with Chemical Suppression of Eluent Conductivity     4110 C. Single-Column Ion Chromatography with Direct Conductivity Detection     4110 D. Ion Chromatographic Determination of Oxyhalides and Bromide   |
| 3. Chlorite                             | 7758-19-2  | 0.7                                  | <ul> <li>4110 B. Ion Chromatography with<br/>Chemical Suppression of Eluent<br/>Conductivity</li> <li>4110 C. Single-Column Ion<br/>Chromatography with Direct<br/>Conductivity Detection</li> <li>4110 D. Ion Chromatographic<br/>Determination of Oxyhalides and<br/>Bromide</li> </ul> |
| 4. Dibromoacetonitrile                  | 3252-43-5  | 0.07                                 | Gas Chromatography / Electron Capture Detector  |

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<sup>&</sup>lt;sup>1</sup>Note: WHO 2011 did not specify any guideline values for chlorine dioxide because of its rapid hydrolysis to chlorite. In addition, the provisional guideline value for chlorite (i.e. 0.7 mg/L) is considered to be adequately protective against potential toxicity from chlorine dioxide. The taste and odor threshold for this compound is 0.4 mg/L [4]

| Parameter                      | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level (mg/L) | Method of Analysis   |
|--------------------------------|--|--------------------------------------|--|
| 5. Dichloroacetate             | 79-43-6  | 0.05                                 | <ul> <li>Gas Chromatography / Electron<br/>Capture Detector</li> <li>Gas Chromatography / Mass<br/>Spectrometry</li> <li>6251 B. Micro Liquid-Liquid<br/>Extraction Gas Chromatographic<br/>Method</li> </ul>  |
| 6. Dichloroacetonitrile        | 3018-12-0  | 0.02                                 | <ul> <li>Gas Chromatography / Electron<br/>Capture Detector</li> <li>5710 D. Formation of Other<br/>Disinfection By-Products (DBPs)</li> <li>USEPA 551.1</li> </ul>  |
| 7. Monochloroacetate           | 79-11-8  | 0.02                                 | 6251 B. Micro Liquid-Liquid<br>Extraction Gas Chromatographic<br>Method  |
| 8. Trichloroacetate            | 76-03-9  | 0.2                                  | 6251 B. Micro Liquid-Liquid     Extraction Gas Chromatographic     Method  |
| 9. 2,4,6-Trichlorophenol       | 88-06-2  | 0.2                                  | <ul> <li>6251 B. Micro Liquid-Liquid Extraction Gas Chromatographic Method</li> <li>6410 B. Liquid-Liquid Extraction Gas Chromatographic/Mass Spectrometric Method</li> <li>6420 B. Liquid-Liquid Extraction Gas Chromatographic Method</li> </ul>   |
| Trihalomethanes                |  |                                      | emematograpme 1.150nou   |
| 10. Bromoform                  | 75-25-2  | 0.1                                  | <ul> <li>6040 B. Closed-Loop Stripping, Gas Chromatographic/Mass Spectrometric Analysis;</li> <li>6200 B. Purge and Trap Capillary-Column Gas Chromatographic/Mass Spectrometric Method</li> <li>6200 C. Purge and Trap Capillary – Column Gas Chromatographic Method</li> <li>6232 B. Liquid-Liquid Extraction Gas Chromatographic Method</li> </ul>                              |
| 11.Bromodichloromethane (BDCM) | 75-27-4  | 0.06                                 | <ul> <li>6040 B. Closed-Loop Stripping, Gas<br/>Chromatographic/Mass Spectrometric<br/>Analysis;</li> <li>6200 B. Purge and Trap Capillary-<br/>Column Gas Chromatographic/Mass<br/>Spectrometric Method</li> <li>6200 C. Purge and Trap Capillary –<br/>Column Gas Chromatographic<br/>Method</li> <li>6232 B. Liquid-Liquid Extraction Gas<br/>Chromatographic Method</li> </ul> |
| 12. Chloroform                 | 67-66-3  | 0.3                                  | <ul> <li>6200 B. Purge and Trap Capillary-<br/>Column Gas Chromatographic/Mass<br/>Spectrometric Method</li> <li>6200 C. Purge and Trap Capillary –<br/>Column Gas Chromatographic<br/>Method</li> </ul>   |

| Parameter                      | Chemical<br>Abstracts<br>Service<br>(CAS)<br>No. | Maximum<br>Allowable<br>Level (mg/L) | Method of Analysis   |
|--------------------------------|--|--------------------------------------|--|
|                                |  |                                      | 6232 B. Liquid-Liquid Extraction Gas<br>Chromatographic Method   |
| 13.Dibromochloromethane (DBCM) | 124-48-1   | 0.1                                  | <ul> <li>6040 B. Closed-Loop Stripping, Gas<br/>Chromatographic/Mass Spectrometric<br/>Analysis;</li> <li>6200 B. Purge and Trap Capillary-<br/>Column Gas Chromatographic/Mass<br/>Spectrometric Method</li> <li>6200 C. Purge and Trap Capillary –<br/>Column Gas Chromatographic<br/>Method</li> <li>6232 B. Liquid-Liquid Extraction Gas<br/>Chromatographic Method</li> </ul> |
| 14. Total THM                  | N/A  | 1                                    | The sum of the ratio of the concentration of each to its maximum allowable level should not exceed 1.  |

Table A-7. Standard Values and Methods of Analysis for Radiological Parameters

| Radionuclides Analysis                       | Screening Level   | Methods of Analysis   |
|--|---|---|
| 1. Gross alpha                               | 0.5 Bq/L  | <ul> <li>7110 B. Evaporation Method for Gross<br/>Alpha-Beta</li> <li>7110 C. Co-precipitation Method for<br/>Gross Alpha Radioactivity in Drinking<br/>Water</li> <li>Low Level Liquid Scintillation Counting</li> </ul> |
| 2. Gross beta                                | 1.0 Bq/L  | <ul> <li>7110 B. Evaporation Method for Gross<br/>Alpha-Beta</li> <li>7110 C. Co-precipitation Method for<br/>Gross Alpha Radioactivity in Drinking<br/>Water</li> <li>Low Level Liquid Scintillation Counting</li> </ul> |
| 3. Radon                                     | 11.0 Bq/L<br>MCL-maximum<br>contaminant level<br>[EPA 2000] | • 7500-Rn B. Liquid Scintillation Method  |
|  | Guidance Level  |   |
| 4. Gamma* (Ra-226)                           | 1 Bq/L  | • 7120 B. Gamma Spectroscopic Method  |
| 5. Gamma* (Ra-228)                           | 0.1 Bq/L  | • 7120 B. Gamma Spectroscopic Method  |
| 6. Gamma* (Sr-90, I-<br>131, Cs-134, Cs-137) | 10 Bq/L   | 7120 B. Gamma Spectroscopic Method  |
| 7. Tritium* (H-3)                            | 10,000 Bq/L   | • 7500-3H B. Liquid Scintillation Spectrometric Method  |

<sup>\*</sup>Gamma and Tritium in drinking water are analyzed only during emergency situations such as nuclear accidents and radioactive material spills and leakages.

Annex B

**Table B-1. Mandatory Drinking-Water Quality Parameters** 

| No. | Parameter                   | Sampling Location*            |
|-----|-----------------------------|-------------------------------|
| 1   | Thermotolerant Coliform     | Treatment Plant Outlet/Source |
|     | E. coli                     | and Consumers' Taps           |
| 2   | Arsenic (As)                | Treatment Plant Outlet/Source |
| 3   | Cadmium (Cd)                | Consumers' Taps               |
| 4   | Lead (Pb)                   | Consumers' Taps               |
| 5   | Nitrate (NO <sub>3</sub> -) | Treatment Plant Outlet/Source |
| 6   | Color (Apparent)            | Treatment Plant Outlet/Source |
|     |                             | and Consumers' Taps           |
| 7   | Turbidity                   | Consumers' Taps               |
| 8   | pН                          | Treatment Plant Outlet/Source |
|     |                             | and Consumers' Taps           |
| 9   | Total Dissolved Solids      | Treatment Plant Outlet/Source |
| 10  | Disinfectant Residual       | Treatment Plant Outlet/Source |
|     |                             | and Consumers' Taps           |

<sup>\*</sup>applicable to all Level II and Level III water facilities

**Table B-2. Primary Drinking-Water Quality Parameters** 

| No. | Parameter                          | No. | Parameter                             |
|-----|------------------------------------|-----|---------------------------------------|
| 1   | 1,2-Dibromo-3-chloropropane (DBCP) | 29  | Dibromochloromethane (DBCM)           |
| 2   | 1,2-Dichlorobenzene                | 30  | Dibromoacetonitrile                   |
| 3   | 1,2-Dichloroethane                 | 31  | Dichloroacetate                       |
| 4   | 1,2-Dichloroethene                 | 32  | Dichloroacetonitrile                  |
| 5   | 1,4-Dichlorobenzene                | 33  | Dichlorodiphenyltrichloroethane (DDT) |
| 6   | 2,4,6-Trichlorophenol              | 34  | Dichloromethane                       |
| 7   | Acrylamide                         | 35  | Endrin                                |
| 8   | Aldrin and Dieldrin                | 36  | Epichlorohydrin                       |
| 9   | Alpha Particles                    | 37  | Ethylbenzene                          |
| 10  | Atrazine                           | 38  | Ethylene Dibromide                    |
| 11  | Antimony                           | 39  | Fluoride                              |
| 12  | Barium                             | 40  | Glyphosate                            |
| 13  | Benzene                            | 41  | Lindane                               |
| 14  | Benzo(a)pyrene (PAHs)              | 42  | Manganese                             |
| 15  | Beta Particles                     | 43  | Mercury (Total)                       |
| 16  | Boron                              | 44  | Monochloroacetate                     |
| 17  | Bromate                            | 45  | Nickel                                |
| 18  | Bromodichloromethane (BDCM)        | 46  | Nitrite                               |
| 19  | Bromoform                          | 47  | Pendimethalin                         |
| 20  | Carbon Tetrachloride               | 48  | Radon                                 |
| 21  | Carbofuran                         | 49  | Sulfate                               |
| 22  | Chlorate                           | 50  | Selenium                              |
| 23  | Chlordane                          | 51  | Styrene                               |
| 24  | Chlorite                           | 52  | Tetrachloroethene                     |
| 25  | Chloroform                         | 53  | Trichloroacetate                      |
| 26  | Chromium (Total)                   | 54  | Toluene                               |
| 27  | Cyanide (Total)                    | 55  | Total Trihalomethane (THM)            |
| 28  | Di(2-ethylhexyl)phthalate          | 56  | Vinyl chloride                        |

 Table B-3. Secondary Drinking-Water Quality Parameters

| No. | Parameter        | No. | Parameter       |
|-----|------------------|-----|-----------------|
| 1   | Aluminum         | 7   | Odor            |
| 2   | Chloride         | 8   | Sodium          |
| 3   | Copper           | 9   | Taste           |
| 4   | Total Hardness   | 10  | Xylenes (total) |
| 5   | Hydrogen Sulfide | 11  | Zinc            |
| 6   | Iron             |     |                 |

Annex C
Table C-1. Minimum Frequency of Sampling for Microbiological Examination of Drinking-Water

| Source and Mode<br>of Supply                        | Population<br>Served | Minimum Frequency of Sampling for<br>Total Coliform and Thermotolerant<br>coliform/E.coli*   | Minimum Frequency of Sampling for Heterotrophic Plate Count (HPC)*    | Point of<br>Compliance |
|---|----------------------|--|---|------------------------|
| 1. Level I  | -                    | 1 sample every three (3) months  | Not required  | Point source           |
| 2. Level II   | -                    | 1 sample every other month   | 1 sample every other<br>month<br>(required if treated)                | Communal faucet        |
| 3. Level III  | Less than 5,000      | 2 samples monthly  | 2 samples monthly   | Consumer's tap         |
|   | 5,000 –<br>100,000   | 1 sample per 5,000 population + 2 additional samples monthly   | 1 sample per 5,000<br>population + 2<br>additional samples<br>monthly | Consumer's tap         |
|   | More than 100,000    | 1 sample per 10,000 population, plus 12 additional samples monthly  Collection of samples should be spread out within a month  Compliance to total coliform: At least 95% of standard samples taken in each month from each reservoir and distribution point is total coliform negative, provided that thermotolerant coliform is absent  Compliance to thermotolerant coliform: No samples should test positive for thermotolerant coliform | Required, at least<br>40% of the sampling<br>points                   | Consumer's tap         |
| 4. All buildings (i.e. residential, commercial, 600 |                      | 1 sample every other month   | 1 sample every other month  | Consumer's tap         |
| industrial and<br>institutional<br>buildings)       | More than 600        | 1 sample monthly   | Once a month  | Consumer's tap         |
| 5. Food<br>Establishments                           | -                    | 1 sample every other month   | 1 sample every other month  | Consumer's tap         |
| 6. Ice Plants                                       | -                    | Once a month   | Once a month  | Product ice            |

<sup>\*</sup> Refer to Table 1 for specific microbiological point of compliance

Table C-2. Minimum Frequency of Sampling for Mandatory Physical and Chemical Parameters

| ;  | Source and Mode of Supply  | Population<br>Served Per<br>Supply System | Number of<br>Samples/Frequency of<br>Sampling   | Sampling Location  |
|--|--|---|---|--------------------|
| 1.   | Level I (Point Source)   | -   | One sample per year   | Point Source       |
| 2.   | Level II (Communal<br>Faucet)  | -   | One sample per year   | Refer to Table B-1 |
| 3.   | Level III  | 49,999 and below                          | One sample per year   |                    |
|  |  | 50,000 and above                          | One sample for every 250,000 population served per year   |                    |
| <ul><li>4.</li><li>5.</li><li>6. ]</li></ul> | All buildings (i.e. residential, industrial, commercial, and institutional buildings)  Food Establishments  Ice Plants | -   | One sample per year  Parameters to be tested:  • <u>Water from main utilities:</u> Lead, color, odor, turbidity, pH, TDS  • <u>With own source of water:</u> All mandatory parameters  • <u>Water from main utilities</u> | Consumers' taps    |
| o. ree rames                                 |  |   | and own source: All mandatory parameters  |                    |

Table C-3. Minimum Frequency of Sampling for Radiological Parameters

| Туре     | Frequency                                  |
|----------|--|
| Initial  | Four (4) consecutive quarters for one year |
| Periodic | Once every three (3) years                 |

Table C-4. Minimum Frequency of Sampling for Mandatory Microbiological and Physico-Chemical Parameters for Other Modes of Distribution of Drinking-Water

| Source                        | Parameter   | Minimum Frequency of<br>Sampling |
|-------------------------------|---|----------------------------------|
| Water Refilling               | Total Coliform                                    | One sample per month             |
| Station and                   | Thermotolerant coliform/ <i>E. coli</i>           | One sample per month             |
| Water Vending                 | Heterotrophic Plate Count                         | One sample per month             |
| Machines                      | All mandatory physico-<br>chemical parameters     | Two samples per year             |
| Mobile Water<br>Tank and Bulk | Microbiological (total coliform, E. coli and HPC) | Once a month                     |
| Water Supply                  | All mandatory physico-<br>chemical                | Once a year                      |
| Water Refilling               | Other parameters identified by                    | One sample per year or as may be |
| Station, Water                | the LDWQMC  | required by the LDWQMC           |
| Vending                       |   |                                  |
| Machines,                     |   |                                  |
| Mobile Water                  |   |                                  |
| Tank, Bulk                    |   |                                  |
| Water Supply                  |   |                                  |

# Annex D

**Table D-1. Summary of Sampling Requirements for Inorganic Parameters** 

| Parameters             | Container Material  | Minimum Volume<br>of Sample                       | Mode of<br>Preservation  | Holding<br>Time |
|------------------------|---|---|--|-----------------|
| 1. Antimony            | Plastic/Polyethylene<br>or Glass containers<br>rinsed with 50%<br>HNO <sub>3</sub>  | 100 mL  | Add HNO <sub>3</sub> to pH < 2   | 28 days         |
| 2. Arsenic             | Plastic/Polyethylene<br>or Glass containers<br>rinsed with 50%<br>HNO <sub>3</sub>  | 100 mL  | Add HNO <sub>3</sub> to pH < 2   | 28 days         |
| 3. Barium              | Plastic/Polyethylene<br>or Glass containers<br>rinsed with 50%<br>HNO <sub>3</sub>  | 100 mL  | Add HNO <sub>3</sub> to pH < 2   | 28 days         |
| 4. Boron               | Polyethylene bottles or alkali-resistant, boron-free glassware  Fill container completely to exclude air  | 100 mL (Fill container completely to exclude air) | Store all reagents in<br>polyethylene or<br>boron-free<br>containers             | 28 days         |
| 5. Cadmium             | Plastic/Polyethylene<br>or Glass containers<br>rinsed with 50%<br>HNO <sub>3</sub>  | 100 mL  | Add HNO <sub>3</sub> to pH < 2   | 28 days         |
| 6. Chromium<br>(Total) | Plastic/Polyethylene<br>or Glass containers<br>rinsed with 50%<br>HNO <sub>3</sub>  | 100 mL  | Add HNO <sub>3</sub> to pH < 2   | 28 days         |
| 7. Cyanide (Total)     | Dark<br>Polyethylene/Plastic<br>or Glass bottle   | 500 mL  | Add NaOH to pH>12. Remove sulfide Refrigerate in the dark.                       | 24 hours        |
| 8. Fluoride            | Preferred: Polyethylene bottles  Glass bottles if bottle does not contain high-fluoride solutions Polytetrafluoroethyle ne (PTFE) containers are not suitable | 200 mL  | None required  | 28 days         |
| 9. Lead                | Plastic/Polyethylene<br>or Glass containers<br>rinsed with 50%<br>HNO <sub>3</sub>  | 100 mL  | Add HNO <sub>3</sub> to pH < 2   | 28 days         |
| 10. Manganese          | Acidified<br>Polyethylene bottle  | 1 liter   | Acidify sample at<br>the time of<br>collection with<br>HNO <sub>3</sub> to pH <2 | 28 days         |
| 11. Mercury (Total)    | Glass containers<br>rinsed with 50%<br>HNO <sub>3</sub>   | 500 mL  | Add HNO <sub>3</sub> to unfilteredsample to $pH < 1$ .                           | 28 days         |

| Parameters   | Container Material               | Minimum Volume<br>of Sample | Mode of<br>Preservation                           | Holding<br>Time         |
|--------------|----------------------------------|-----------------------------|---|-------------------------|
|              |                                  |                             | Add K <sub>2</sub> Cr <sub>2</sub> O <sub>3</sub> |                         |
| 12. Nickel   | Plastic or Glass bottle          | 500 mL                      | Add HNO <sub>3</sub> to pH <2                     | 6 months                |
| 13. Nitrate  | Glass or<br>Plastic/Polyethylene | 500 mL                      | Refrigerate (unfiltered samples)                  | 24 hours                |
|              | container                        |                             | Filter on site                                    | 1 month –               |
|              |                                  |                             | (0.45 m cellulose                                 | consult                 |
|              |                                  |                             | acetate membrane                                  | analyst                 |
|              |                                  |                             | filter and freeze)                                | depending on analytical |
|              |                                  |                             |   | method                  |
| 14. Nitrite  | Glass or                         | Colorimetric Method:        | Freeze at -20°C or                                | 1 to 2 days             |
|              | Plastic/Polyethylene container   | 50 mL sample                | store at 4°C.                                     |                         |
| 15. Selenium | Plastic/Polyethylene             | 100 mL                      | Add HNO <sub>3</sub> to pH <                      | 28 days                 |
|              | or Glass containers              |                             | 2   |                         |
|              | rinsed with 50%                  |                             |   |                         |
|              | $HNO_3$                          |                             |   |                         |

Sources: APHA 22nd ed., 2007 PNSDW, 2011 ADWG

**Table D-2. Summary of Sampling Requirements for Organic Parameters** 

| Parameters               | Container<br>Material   | Minimum<br>Volume of<br>Sample | Mode of Preservation  | Holding<br>Time  |  |  |
|--------------------------|---|--------------------------------|---|--|--|--|
|                          | Indust  | rial Pollutants                | 5   |  |  |  |
| 1. Benzene               | Screw-cap vial with a hole in the center and TFE-faced silicone septum            | 2 x 40 mL                      | <ul> <li>Cool, ≤6°C</li> <li>For samples that contain volatile constituents but do not contain residual chlorine, add HCl to pH</li> <li>&lt;2.0(4 drops, 1:1 HCl)</li> <li>For samples that contain residual chlorine, add 1000 mg ascorbic acid/L or 0.008% sodium thiosulfate</li> </ul>                   | 14 days  |  |  |
| 2. Benzo(a)pyrene        | Amber glass<br>bottles with a<br>screw cap<br>lined with<br>TFE                   | 1 liter                        | Cool, ≤6°C     For samples that contain residual chlorine, add 1000 mg ascorbic acid/L or 0.008% sodium thiosulfate   | 7 days until<br>extraction; 40<br>days after<br>extraction |  |  |
| 3. Carbon Tetrachloride  | See Benzene   |                                |   |  |  |  |
| 4. 1,2-Dichlorobenzene   | See Benzene   |                                |   |  |  |  |
| 5. 1,4-Dichlorobenzene   | See Benzene   |                                |   |  |  |  |
| 6. 1,2-Dichloroethane    | See Benzene   |                                |   |  |  |  |
| 7. 1,2-Dichloroethene    | See Benzene   |                                |   |  |  |  |
| 8. Dichloromethane       | See Benzene   | See Benzene                    |   |  |  |  |
| 9. Di(2-ethylhexyl)phtha | alate See Benzo(a)py  | See Benzo(a)pyrene             |   |  |  |  |
| 10. Ethylbenzene         | See Benzene   | See Benzene                    |   |  |  |  |
| 11. Styrene              | See Benzene   |                                |   |  |  |  |
| 12. Tetrachloroethene    | See Benzene   |                                |   |  |  |  |
| 13. Toluene              | See Benzene   |                                |   |  |  |  |
| 14. Vinyl Chloride       | See Benzene   |                                |   |  |  |  |
| 15. Xylenes (total)      | See Benzene   |                                |   |  |  |  |
| Pesticides               |   |                                |   |  |  |  |
| Aldrin and Dieldrin      | See Benzo(a)py  | rene                           |   |  |  |  |
|                          | Amber glass bottles filled with a screw cap lined with TFE.                       | 1 liter                        | Cool, ≤6° CFor<br>samples that contain<br>residual chlorine, add<br>1000 mg ascorbic<br>acid/L or 0.008%<br>sodium thiosulfate  | 7 days until<br>extraction; 40<br>days after<br>extraction |  |  |
| 2. Atrazine              | See Aldrin and  |                                |   |  |  |  |
| 3. Carbofuran            | Amber glass bottles fitted with polytetrafluor oethylene (PTFE)-lined screw caps. | None<br>specified              | <ul> <li>Add a sufficient amount of potassium dihydrogen citrate to yield a concentration in the sample of 9.2 to 9.5 g/L to prevent hydrolysis of oxamyl, 3-hydroxycarbofuran, carbaryl, and methiocarb</li> <li>Add sodium thiosulfate to yield a sample concentration in the range of 80 to 320</li> </ul> | 28 days  |  |  |

| Parameters                                | Container<br>Material   | Minimum<br>Volume of<br>Sample      | Mode of Preservation   | Holding<br>Time   |
|---|---|-------------------------------------|--|---|
|   |   |                                     | mg/L to eliminate residual chlorine During transport: Ice, temperature should not exceed 10 degrees Celsius during the first 48 hours after collection In the laboratory: Store samples at temperature below 6 degrees Celsius and protect from light until extraction. Do not freeze sample |   |
| 4. Chlordane                              | See Benzo(a)py  |                                     |  |   |
| 5. 1,2-Dibromo-3-<br>chloropropane (DBCP) | See Aldrin and See Benzene  |                                     |  |   |
| 6. Dichlorodiphenyltrichloroet hane (DDT) | See Benzo(a)py<br>See Aldrin and                                    |                                     |  |   |
| 7. Endrin                                 | See Benzo(a)py  |                                     |  |   |
| , Didini                                  | See Aldrin and Dieldrin   |                                     |  |   |
| 8. Ethylene Dibromide or 1,2-             | See 1,2-Dibrom  | o-3-chloropro                       | pane (DBCP)  |   |
| Dibromoethane                             | See 1,2-Dichlorobenzene   |                                     |  |   |
|   | See Benzene   | T = 0.0 =                           | 1  | T   |
| 9. Glyphosate                             | Polypropylene or amber glass container                              | 500 mL<br>representat<br>ive sample | Add 100 mg/L sodium<br>thiosulfate for chlorinated<br>water and store at 4°C   | 14 days   |
| 10. Lindane                               | See Aldrin and  | D: 11.1.1.                          | away from light  |   |
| 10. Lindane                               | Disinfection Che  |                                     | v_Products   |   |
| 1. Acrylamide                             | See Benzene   | emicuis unu B                       | y-1 Tourcis  |   |
| 2. Epichlorohydrin                        | See Benzene   |                                     |  |   |
| 3. Chlorine Dioxide                       | None  | None                                | Analyse immediately  | -   |
|   | specified   | specified                           |  |   |
| 4. Chlorine Residual                      | Plastic<br>(polyethylene<br>or equivalent)<br>or Glass<br>container | 500 mL                              | Analyse immediately. Keep out of direct sunlight   | 5 minutes   |
| 5. Bromate                                | Glass or  | 500 mL                              | Refrigerate  | 24 hours  |
|   | Plastic/Polyet<br>hylene<br>container                               |                                     | (unfiltered samples) Filter on site (0.45 µm cellulose acetate membrane filter and freeze)   | 1 month -<br>Consult<br>analyst<br>depending on<br>analytical<br>method |
| 6. Chlorate                               | Glass or  | 500 mL                              | Refrigerate  | 24 hours  |
|   | Plastic/Polyet<br>hylene<br>container                               |                                     | (unfiltered samples) Filter on site (0.45 µm cellulose acetate membrane filter and freeze)   | 1 month -<br>Consult<br>analyst<br>depending on<br>analytical<br>method |
| 7. Chlorite                               | Glass or  | 500 mL                              | Refrigerate  | 24 hours  |
|   | Plastic/Polyet  |                                     | (unfiltered samples)   |   |

| hylene container  hylene (0.45 μm cellulose acetate membrane filter and freeze)  hylene (0.45 μm cellulose acetate manlyst depending or analytical method  hylene (0.45 μm cellulose acetate membrane filter and freeze)  hylene (0.45 μm cellulose acetate membrane filter and freeze)  hylene (0.45 μm cellulose acetate manlyst depending or analytical method  -14 Days  holending hosphate dibasic/99% potassium phosphate monobasic + 0.6% ammonium chloride)  9. Dichloroacetate  Screw-cap vial with a hole in the (depending center and TFE-faced silicone septum; zero headspace  septum; zero headspace  10. Dichloroacetonitrile  Glass vial with TFE-  hylene (0.45 μm cellulose acetate membrane filter and freeze)  -14 Days  -14 Days  -14 Days  -14 Days   |
|---|
| with TFE- lined screw caps  -1 gram/ 60-ml amber glass vial (1% sodium phosphate dibasic/99% potassium phosphate monobasic + 0.6% ammonium chloride)  9. Dichloroacetate  Screw-cap vial with a hole in the center and TFE-faced silicone septum; zero headspace  10. Dichloroacetonitrile  With TFE-    -1 gram/ 60-ml amber glass   -14 days   -17 days   -18 |
| vial with a hole in the center and TFE-faced silicone septum; zero headspace  10. Dichloroacetonitrile  vial with a hole in the (depending on vial used)  TFE-faced silicone septum; zero headspace  10. Dichloroacetonitrile  Vial with a mL (depending on vial used)  11°C  21 days for sample extracts freeze at - 11°C  11°C  Cool, ≤6°C -1 gram/ 60-ml amber glass   |
| with TFE1 gram/ 60-ml amber glass   |
| lined screw caps  vial (1% sodium phosphate dibasic/99% potassium phosphate monobasic + 0.6% ammonium chloride)   |
| 11. Monochloroacetate See Dichloroacetate   |
| 12. Trichloroacetate See Dichloroacetate  |
| 13. 2,4,6-Trichlorophenol See Dichloroacetate   |
| See Benzo(a)pyrene  |
| Amber glass bottles with a screw cap lined with TFE  Amber glass bottles with a screw cap lined with TFE  • Refrigerate at 4°C 40 days • Add 80 g sodium thiosulfate per liter of sample if residual chlorine is present  |
| 14. Bromoform See 1,2-Dichlorobenzene   |
| See Benzene   |
| Glass bottle sealed with TFE-lined screw caps   |
| 15. Bromodichloromethane See 1,2-Dichlorobenzene  |
| (BDCM) See Benzene  |
| See Bromoform   |
| 16. Chloroform See Benzene  |
| See Bromoform   |
|   |
| 17. Dibromochloromethane See 1,2-Dichlorobenzene  |
|   |

Sources: APHA 22nd ed., 2011 ADWG

Table D-3. Summary of Sampling Requirements for Physical and Chemical Parameters for Acceptability Aspects

| Parameter           | Container<br>Material  | Minimum Volume of Sample  | Mode of<br>Preservation  | <b>Holding Time</b>  |
|---------------------|--|---|--|--|
| 1. Taste            | <ul><li>Glass-stoppered bottles</li><li>TFE-lined enclosures</li></ul>                           | 500 mL  | Keep cool at $\leq$ 6 $^{0}$ C   | Not more than 6 hrs  |
| 2. Odor             | <ul><li>Glass-stoppered bottles</li><li>TFE-lined enclosures</li></ul>                           | 500 mL  | Keep cool at ≤ 6 ° C   | 6 hrs  |
| 3. Color            | <ul> <li>Acid-washed<br/>amber glass<br/>bottles</li> <li>Covered plastic<br/>bottles</li> </ul> | 100 mL  | Keep cool<br>Analyze same day  | 24 hours   |
| 4. Turbidity        | <ul><li>Polyethylene bottle</li><li>Glass bottle</li></ul>                                       | 100 mL  | Keep cool at ≤ 4°C   | 24 hrs   |
| 5. Aluminum         | Acid-rinsed Plastic bottles  | 25 mL or a portion<br>diluted to 25 mL<br>(In absence of<br>fluoride and complex<br>phosphates) | Examine sample immediately   | Examine sample immediately   |
| 6. Chloride         | <ul><li>Plastic bottle</li><li>Glass bottle</li></ul>  | 100 mL (maximum<br>sample portion)<br>or a suitable portion<br>diluted to 100 mL                | No special preservative is necessary   | 28 days  |
| 7. Copper           | Acidified<br>Polyethylene Bottle   | 1 liter   | Use 0.5 mL 1 + 1<br>HCl/100mL sample<br>or acidify to pH <2<br>with HNO <sub>3</sub> | 28 days  |
| 8. Total Hardness   | Plastic/Glass<br>Container   | 500 mL  | Add HNO <sub>3</sub> or<br>H <sub>2</sub> SO <sub>4</sub>                            | 7 days   |
| 9. Hydrogen sulfide | Glass bottle   | 100 mL  | Preserve using zinc acetate solution   | <ul> <li>2 weeks for<br/>refrigerated<br/>samples</li> <li>1 month for<br/>frozen<br/>samples</li> </ul> |
| 10. Iron            | Acidified Polyethylene Bottle  | 50 mL   | Use 0.5 mL 1 + 1<br>HCl/100mL sample<br>or acidify to pH <2<br>with HNO <sub>3</sub> | 28 days  |
| 11. pH              | Polyethylene<br>bottles  | 50 ml   | None required  | Analyze<br>immediately or<br>not to exceed 6<br>hours after<br>sample<br>collection                      |

| Parameter                  | Container<br>Material  | Minimum Volume of<br>Sample | Mode of<br>Preservation   | Holding Time                                    |
|----------------------------|--|-----------------------------|---|---|
| 12. Sodium                 | Polyethylene bottles   | 1 liter                     | None required   | 28 days   |
| 13. Sulfate                | <ul><li>Polyethylene bottles</li><li>Glass bottles</li></ul>   | 100 mL                      | Keep cool at 4°C  | 7 days  |
| 14. Total Dissolved Solids | <ul><li>Resistant-glass</li><li>Plastic bottles</li></ul>  | 500 mL                      | Keep cool at 4°C  | 7 days  |
| 15. Zinc                   | <ul> <li>Quartz or TFE containers</li> <li>Polypropylene or linear polyethylene with a polyethylene cap</li> <li>Borosilicate glass</li> </ul> | 50 mL                       | • Acidify with concentrated nitric acid to pH <2 • Refrigerate at 4°C | • 6 months • 5 weeks if sample contains mercury |

Sources: APHA 22nd ed.,2007 PNSDW, 2011 ADWG

Table D-4. Summary of Sampling Requirements for Radiological Parameters

| Parameter  | Container Material                                      | Minimum<br>Volume of<br>Sample | Mode of Preservation  | Holding<br>Time |
|------------|---|--------------------------------|---|-----------------|
| 1. Alpha   | Plastic (polyethylene or equivalent) or Glass container | 1L                             | Concentrated HNO <sub>3</sub> or<br>HCl to pH <2  | 28 days         |
| 2. Beta    | Plastic (polyethylene or equivalent) or Glass container | 1L                             | Concentrated HNO <sub>3</sub> or<br>HCl to pH <2  | 28 days         |
| 3. Radon   | Gas-tight PET bottles                                   | 1L                             | Bottles are to be filled full (up to the brim and no air spaces); no acidification required; samples are to be brought to PNRI lab within the day | 8 days          |
| 4. Gamma   | Plastic container                                       | 2L                             | Concentrated HNO <sub>3</sub> or<br>HCl to pH <2  | 6 months        |
| 5. Tritium | Plastic (polyethylene or equivalent) or Glass container | 1L                             | No preservative   | 6 months        |

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Philippine Association of Water Districts

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#### **International Consultants**

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## Stakeholders

Water Service Providers

Angeles City Water District

Baguio Water District

Balamban Water District

Legazpi City Water District

Legazpi City Water District

Balayan Water District
Baliwag Water District
Bansalan Water District
Batac Water District
Batangas City Water District
Limay Water District
Mambajao Water District
Manila Water Company Inc.
Maynilad Water Services Inc.
Metropolitan Cebu Water District

Boracay Island Water Metro Iloilo Water District

Bogo Water District Metro Kidapawan Water District
Butuan City Water District Metro Naga Water District

Cagayan de Oro City Water District
Camarines Norte Water District
Orani Water District

Catbalogan Water District
Catarman Water District
Compostela Water District
San Rafael Water District
Sanchez Mira Water District

Cebu Water St. Joseph Water Services - Laguna

Clark Water Surigao Metro Water District

Dapitan City Water District
Dasmariñas Water District
Tallsay Water District
Tarlac Water District

Davao City Water District Digos Water District

General Santos City Water District

Laguna Water

Toledo City Water District

Water Refilling Station – Aqua Clean Zamboanga City Water District

Water Laboratories and Suppliers

Brownstone Asia-Tech Inc.

Cebu Aqua Lab, Inc.

Chempro Analytical Services Laboratory

Inc.

CRL Environmental Corp.

**Eminent Water Laboratory Center** 

Environmental Health Lab.

F.A.S.T. Laboratory - Cebu, Cubao

HiAdvance Philippines Inc.

Leads Animal Health and Natural

Solutions

MACH Union Water Laboratory National Reference Laboratory

Optimal Laboratory Inc.

Platinum Research Laboratory Inc.

Sanwad Water Laboratory USC Water Laboratory Zamboanga WaterLab Inc.

Academe

Davao Medical School Foundation, Inc. University of the Immaculate Conception

University of the Philippines Diliman-

College of Engineering

University of the Philippines Los Baños University of Southeastern Philippines

University of Visayas

Professional Organizations and Associations

AHFAT Seafood PCAZ

Dencio's Kamayan

**Grand Menseng Hotel** 

Grand Regal Hotel

Health Care Zamboanga

**Integrated Chemists of the Philippines** 

J.V. Angeles Construction Co.

League of Provinces of the Philippines

LES Consultant

MBG Davao City Ice Plant

Pepsi-Cola Products Philippines, Inc.

Philippine Institute of Pure and Applied Chemistry

Pulo Balagtas Rural Waterworks & Sanitation

Association

Philippine Institute of Chemical Engineers Well Drillers Association of the Philippines Philippine Society of Sanitary Engineers

Government Agencies and Regulators

Batangas City Planning & Development

Office

City Health Offices

Department of Education - Region XI

Department of Public Works and

Highways

Department of Trade and Industry-

Bureau of Philippine Standards

Department of Science and Technology Food and Drug Administration – DOH

Health Facilities and Services Regulatory Bureau -

DOH

Local Water Utilities Administration

Metropolitan Waterworks and Sewerage System-

Regulatory Office

National Water Resources Board

Provincial Health Offices